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# Cytomechanic Regulation of Pollen Tube Growth

## MecanX – Understanding Physics of Plant Growth



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Plant Science Center

Amphacademy, Root, September 15<sup>th</sup> 2017



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Eidgenössische Technische Hochschule Zürich  
Swiss Federal Institute of Technology Zurich

Institute for Building Materials

Felix Beyeler

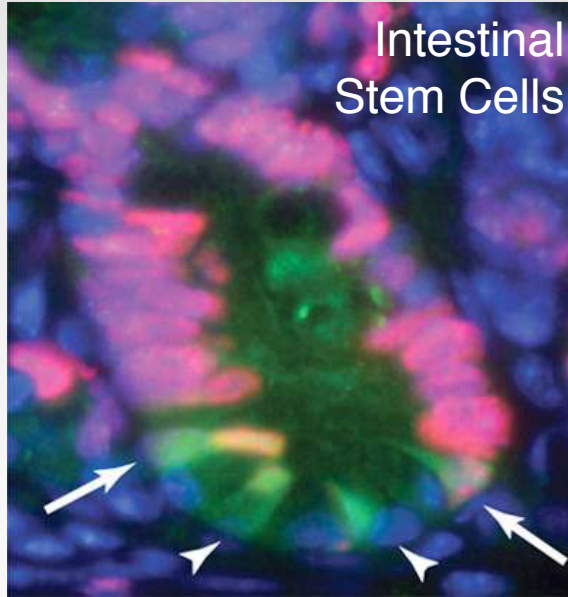


Abu Sebastian



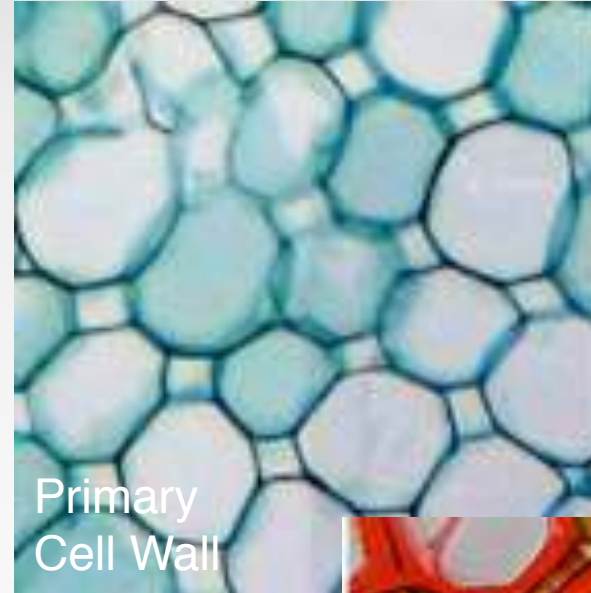
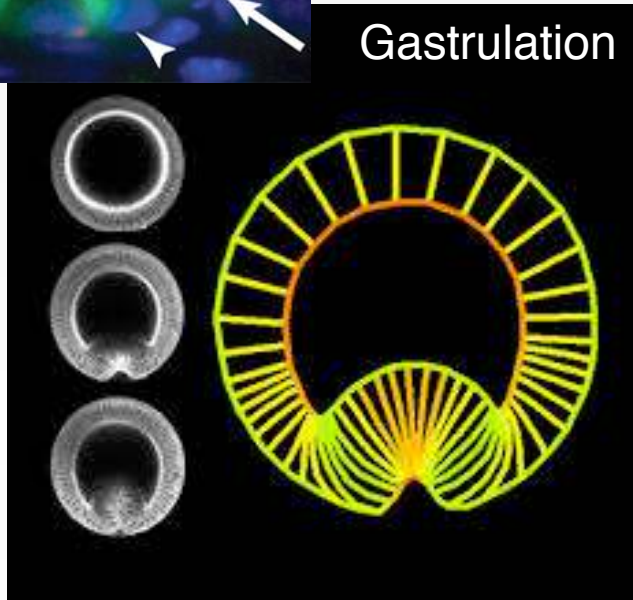
Amphacademy, Root, September 15<sup>th</sup> 2017

# Mechanical Forces Play an Important Role in Morphogenesis

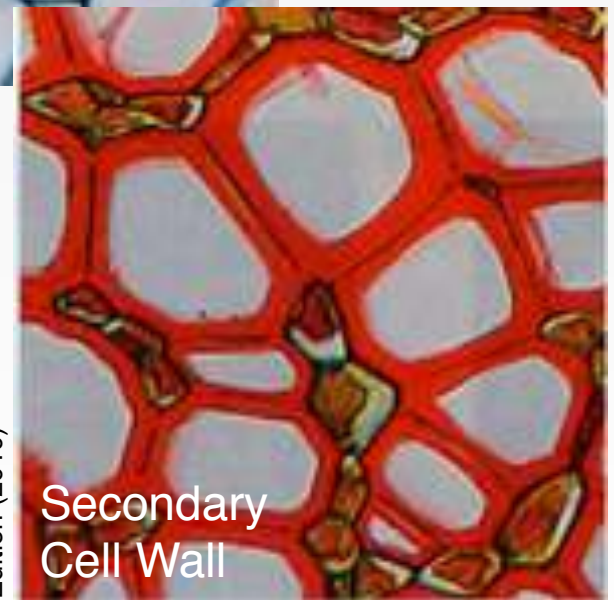


Gastrulation

Broadland et al. (2010) *PNAS* 107, 22111  
 van Dussen et al. (2011) *Development* 139, 488



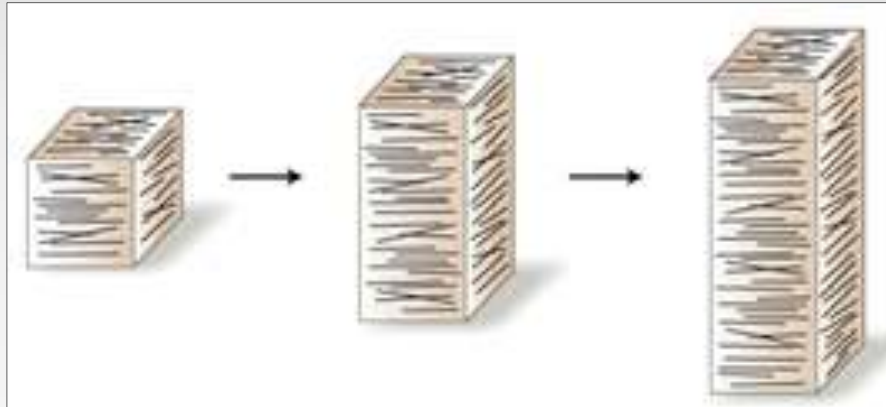
Taiz and Zeiger, *Plant Physiology* 5th Edition (2010)



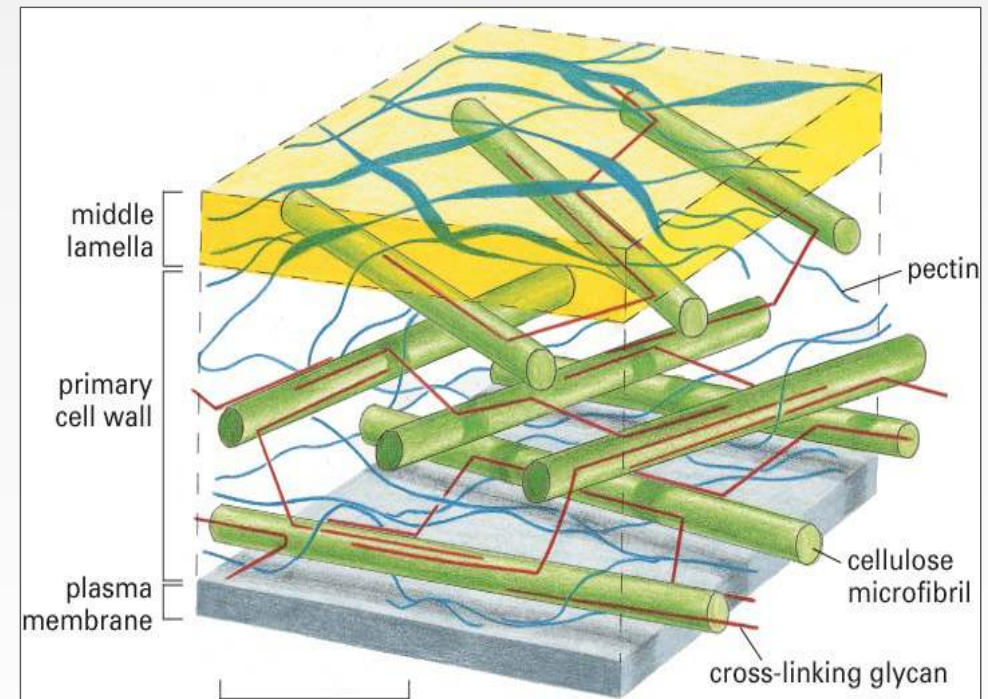


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# Cell Wall and Plant Growth



- cell expansion is driven by **turgor pressure**
- requires coordination between **loosening/deformation** of existing cell wall and the secretion of new cell wall material
- relationship between **biochemical** composition and **mechanical** properties of the cell wall are **not well understood**



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# Mechanics of Pollen Tube Growth



Movie: Peter Hepler

Goals of the **MecanX** project are:

- to develop novel methods to **measure** mechanophysical properties
- to investigate how the biochemical composition affects the **mechanical properties** of the cell wall
- to develop **mathematical models** to describe plant cell growth based on cytomechanical parameters

We use **pollen tubes** as our model system because:

- they are **single** cells (no confounding effects from neighboring cells)
- cell expansion is restricted to the **tip** (2-D system)
- grow very fast, such that mechanical changes are **exaggerated** and can be followed more easily



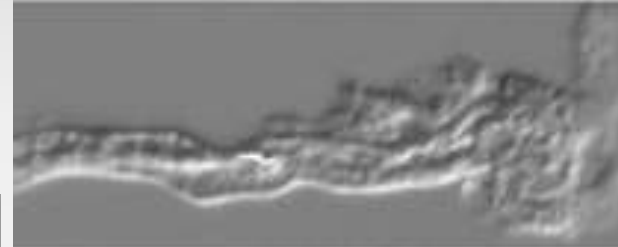
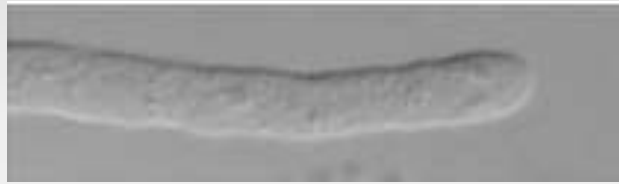
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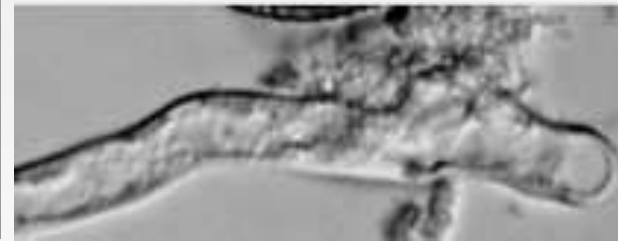
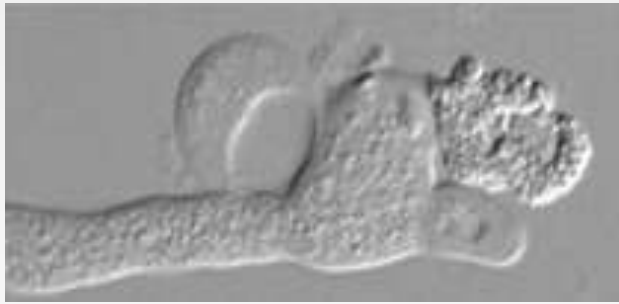
# Mutants Affecting Cell Wall Composition of the Pollen Tube

wild-type

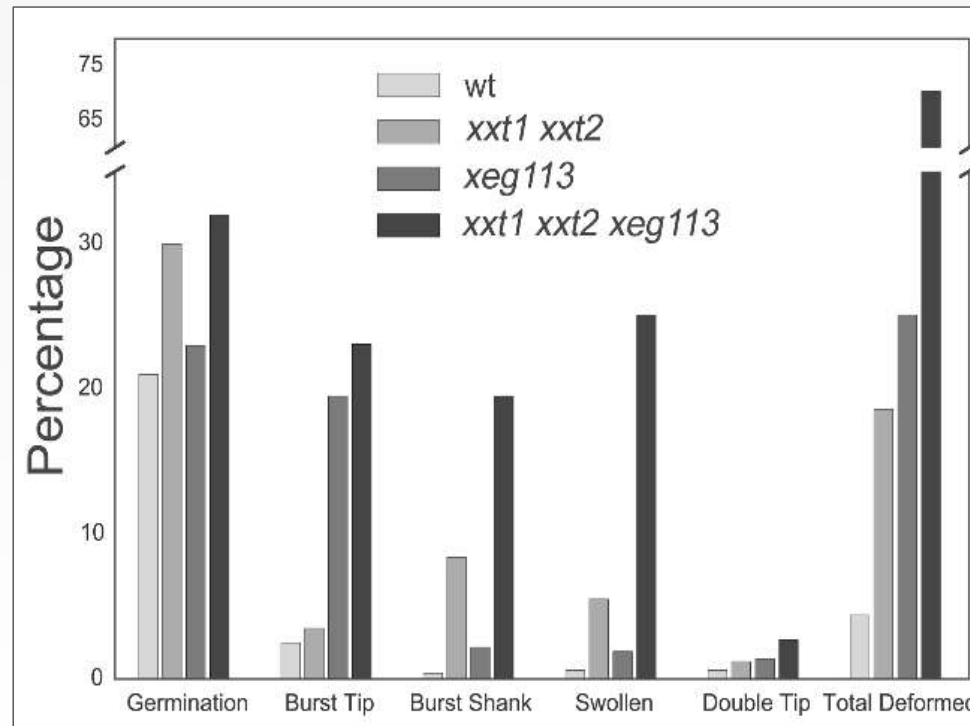


*xeg113*  
(non-functional  
extensins)

*xxt1 xxt2*  
(deficient in  
xyloglucans)



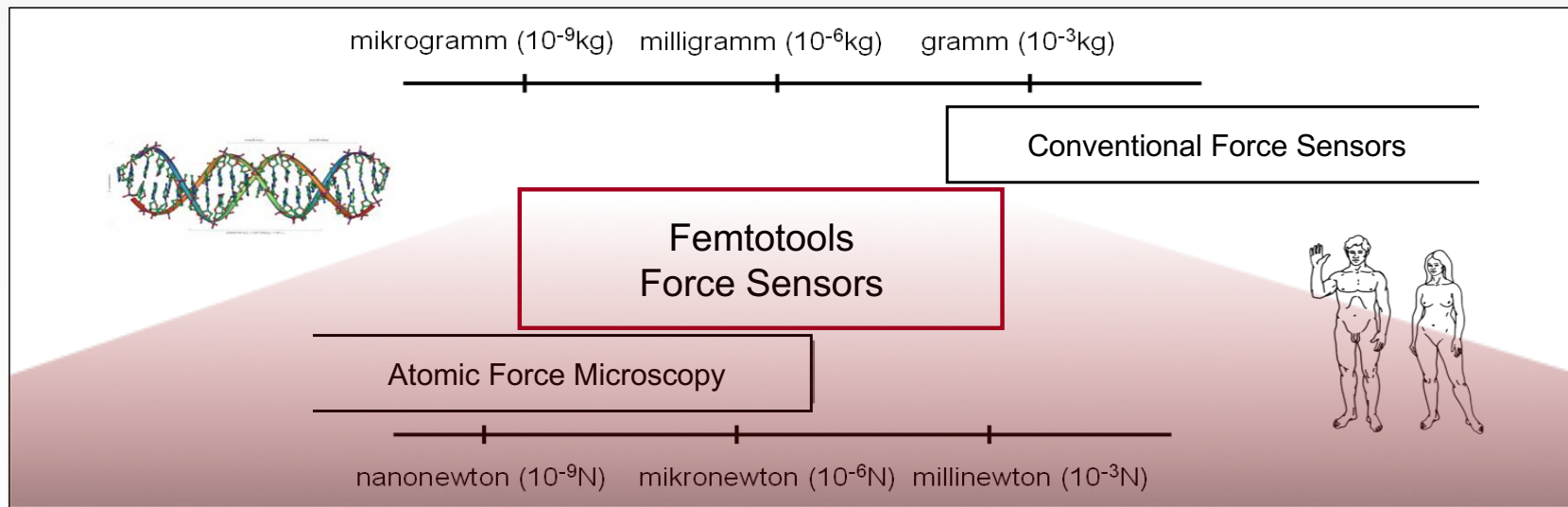
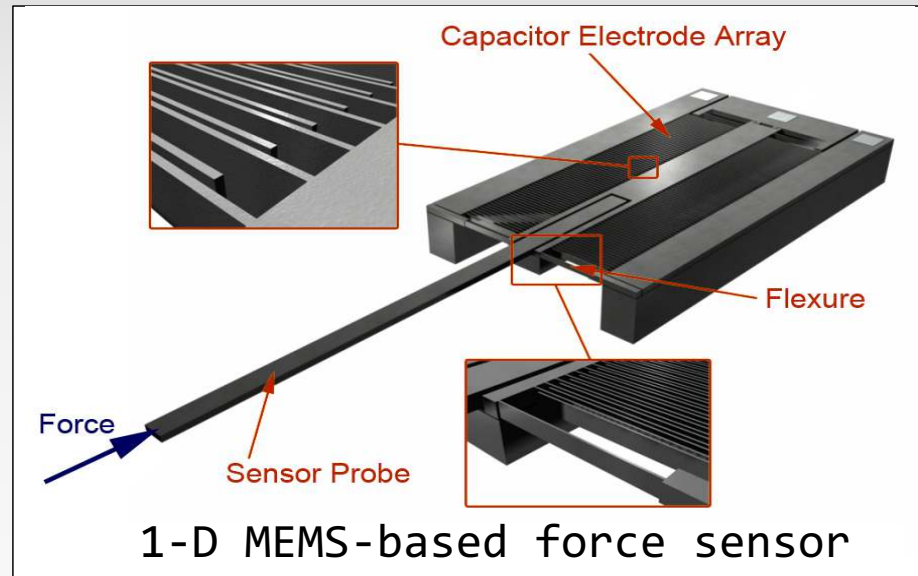
*xxt1 xxt2*  
*xeg113*





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# Microelectronic Mechanical Systems (MEMS)-Based Force Sensor



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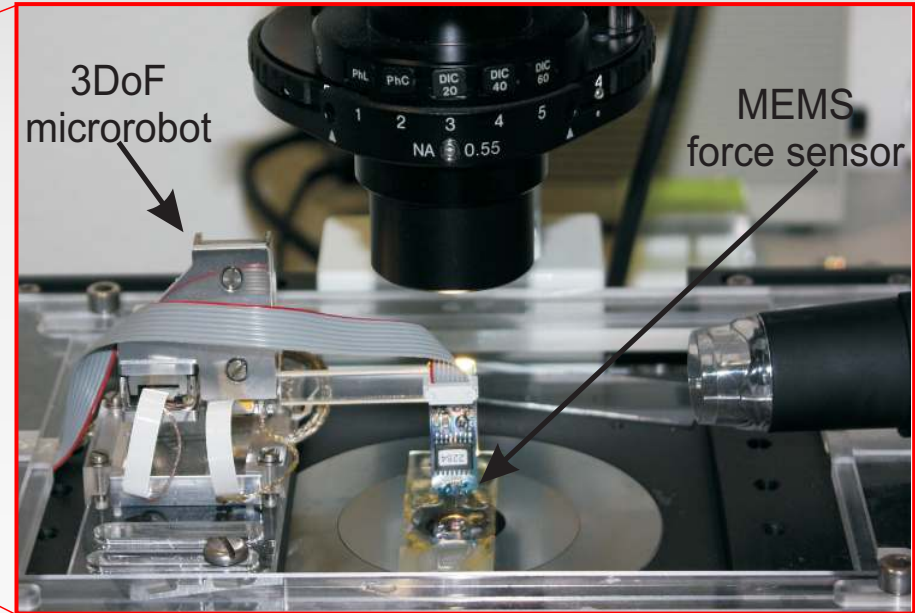
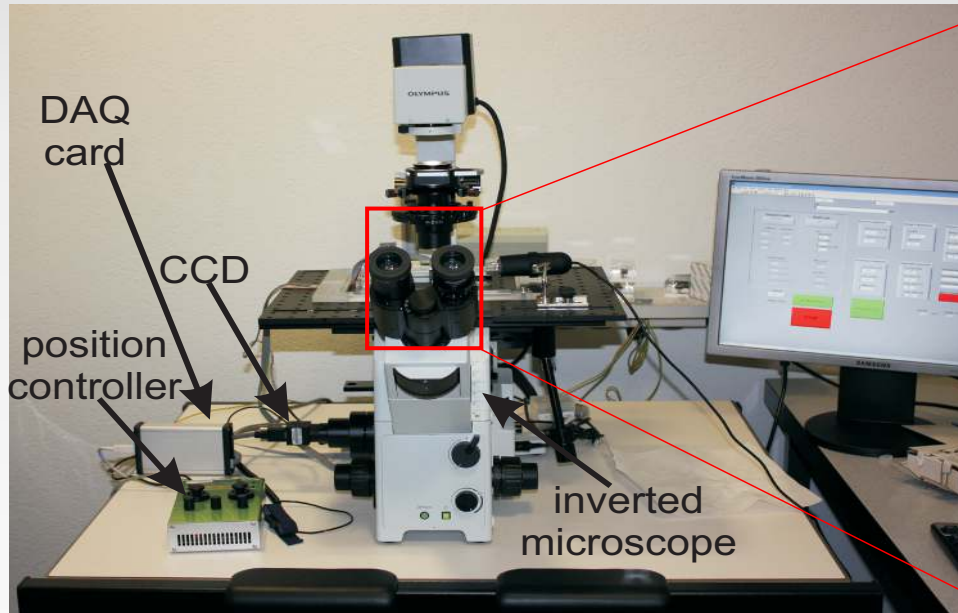
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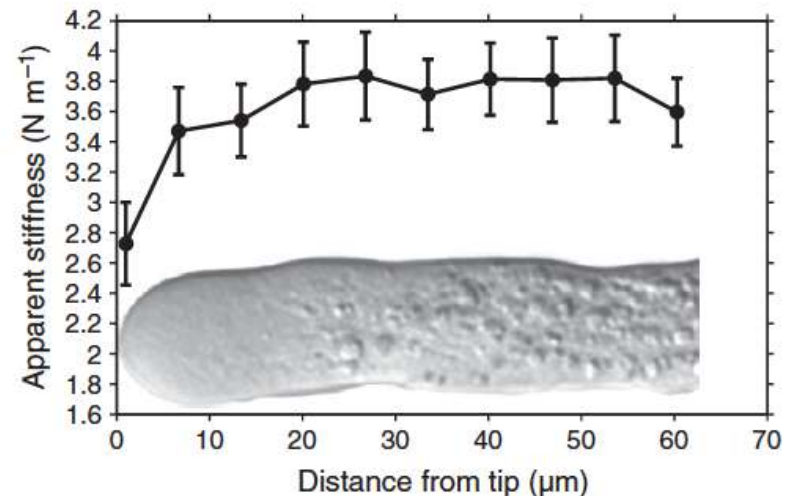
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# Cellular Force Microscope (CFM)



Measured **apparent stiffness** depends on:

- stiffness of the cell wall (Young's modulus and wall thickness)
- turgor pressure
- geometry of the cell and interaction with the force sensor



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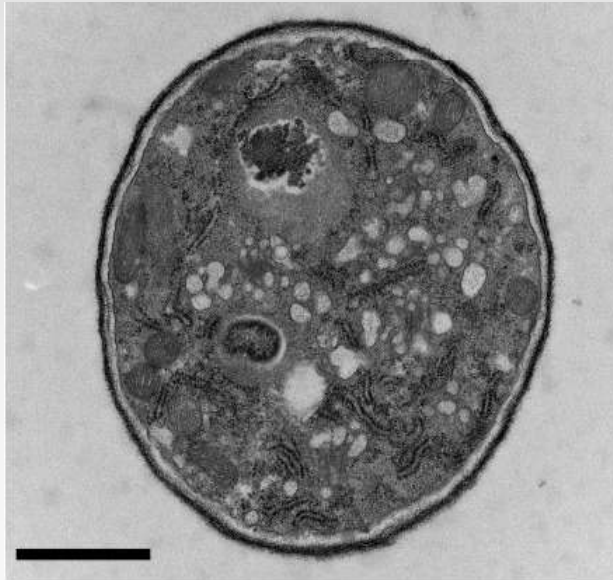
Vogler et al. (2013) *Plant J.* 73, 617  
Felekis et al (2011) *Micro & Nano Lett.* 6, 311





# Measuring Cell Wall Thickness by TEM

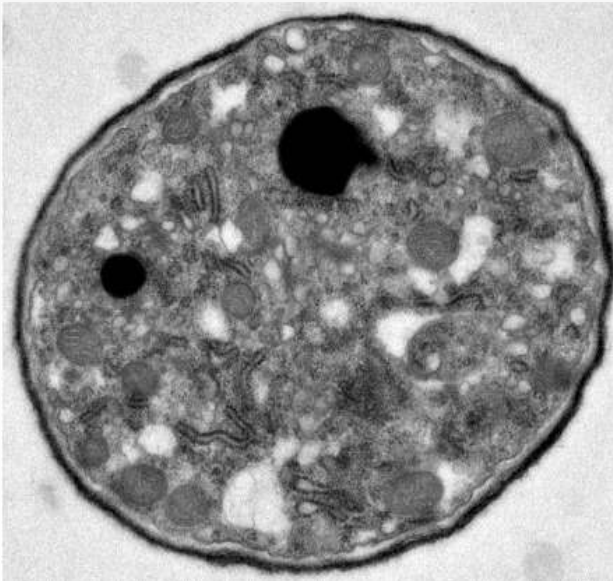
wild-type



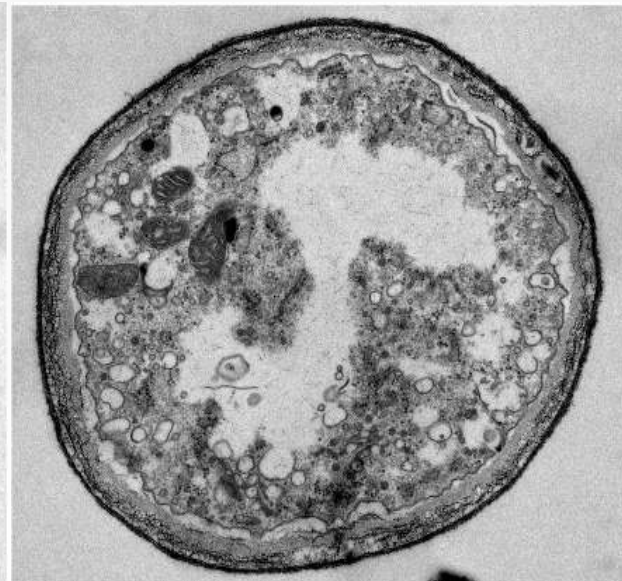
*xxt1 xxt2*



*xeg113*



*xxt1 xxt2*  
*xeg113*

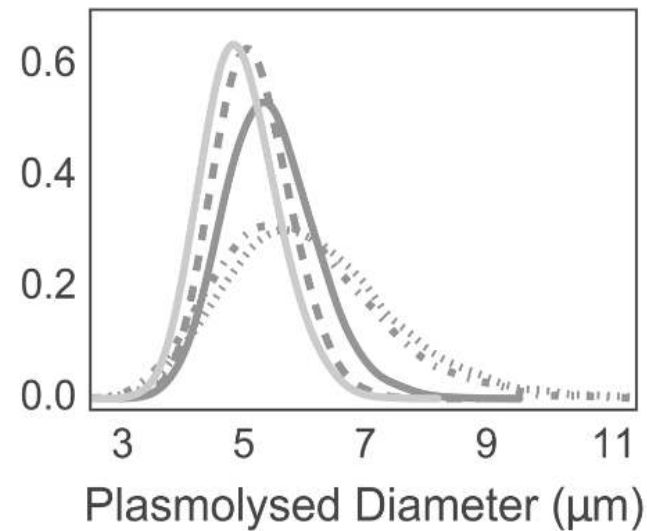
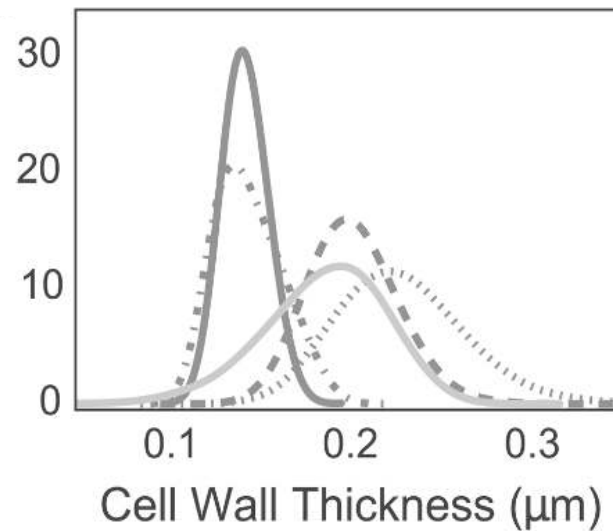
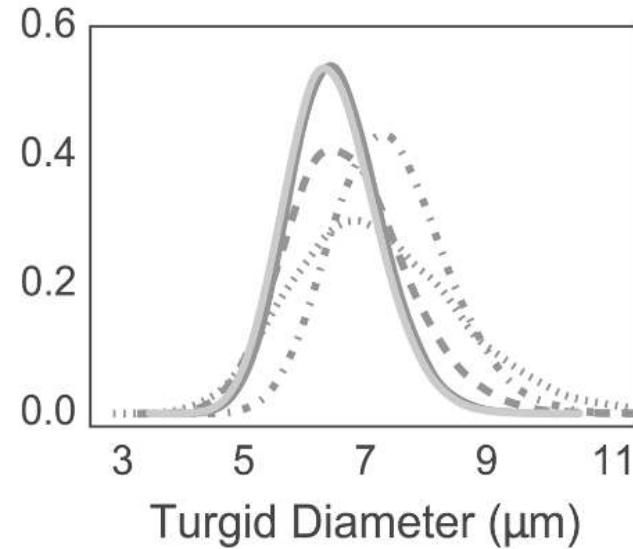
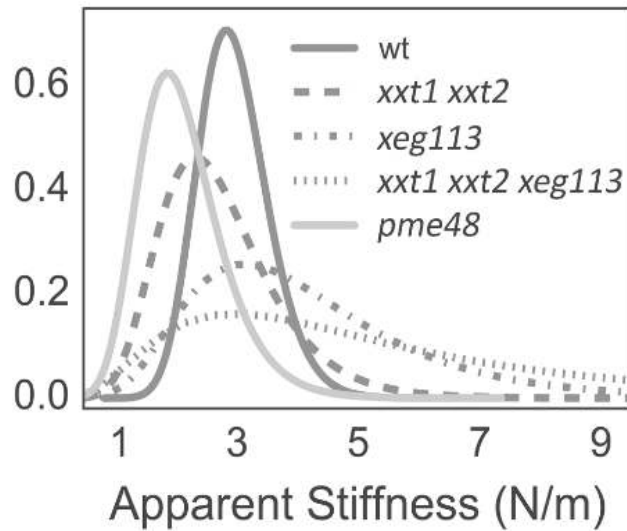




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# Probability Density Functions of Input Parameters for FEM Modeling

Probability Density Function

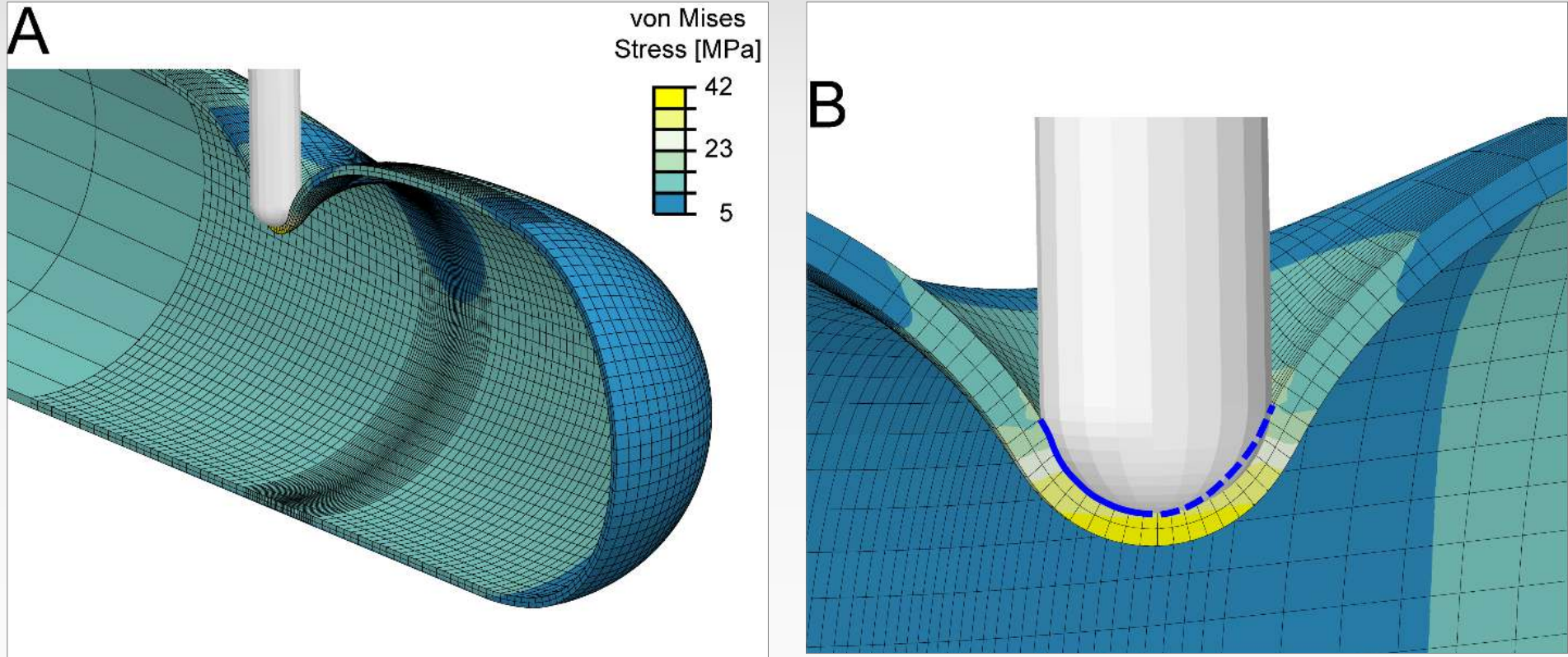


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Gautam Munglani, Hannes Vogler

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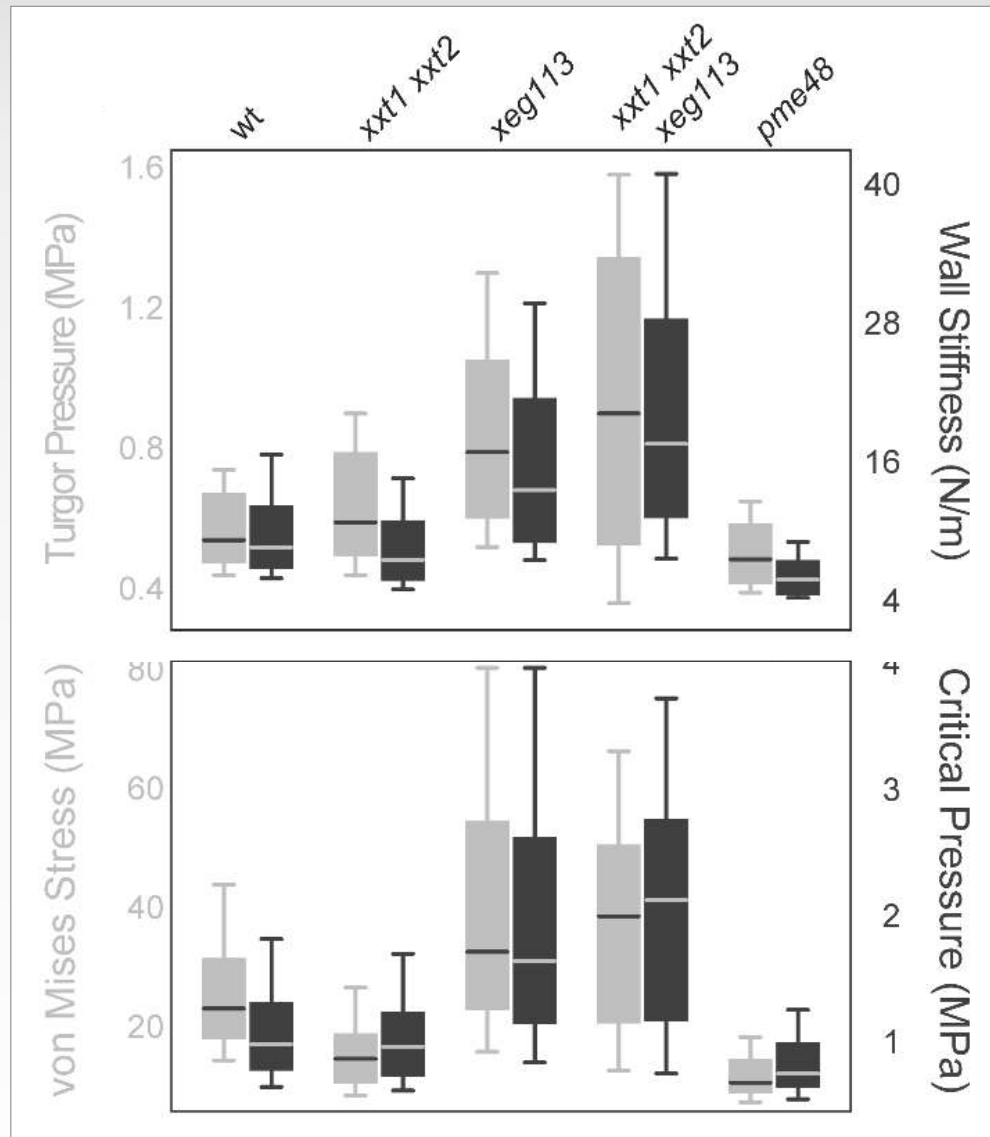


Three-step FEM-based model that sequentially fits the input parameters:

- cell wall thickness (TEM)
- circumferential stretch (ratio of turgid to plasmolyzed PT diameter)
- apparent stiffness (CFM)



# FEM-Based Modeling to Extract Turgor and Young's Modulus



Xyloglucan-deficient PTs have a slightly elevated median turgor pressure and a reduced median cell wall stiffness.

Extensin-deficient and triple mutant PTs have a significantly increased turgor pressure and wall stiffness compared with much higher variance. Turgor and wall stiffness show a synergistic relationship.

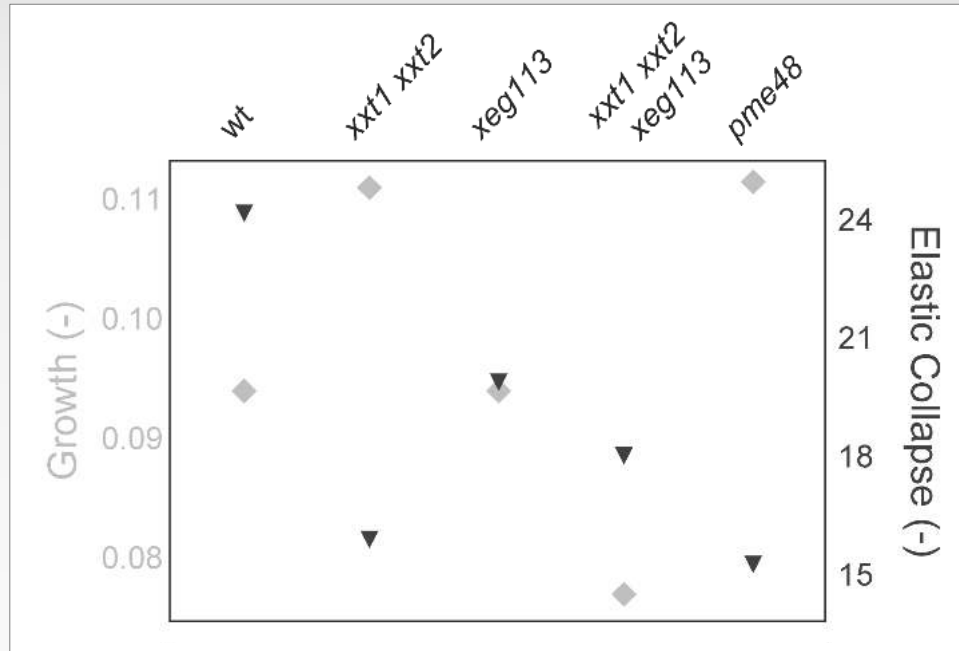
Modeling von Mises stresses and critical pressure shows similar changes consistent with PT bursting frequencies in the mutants.





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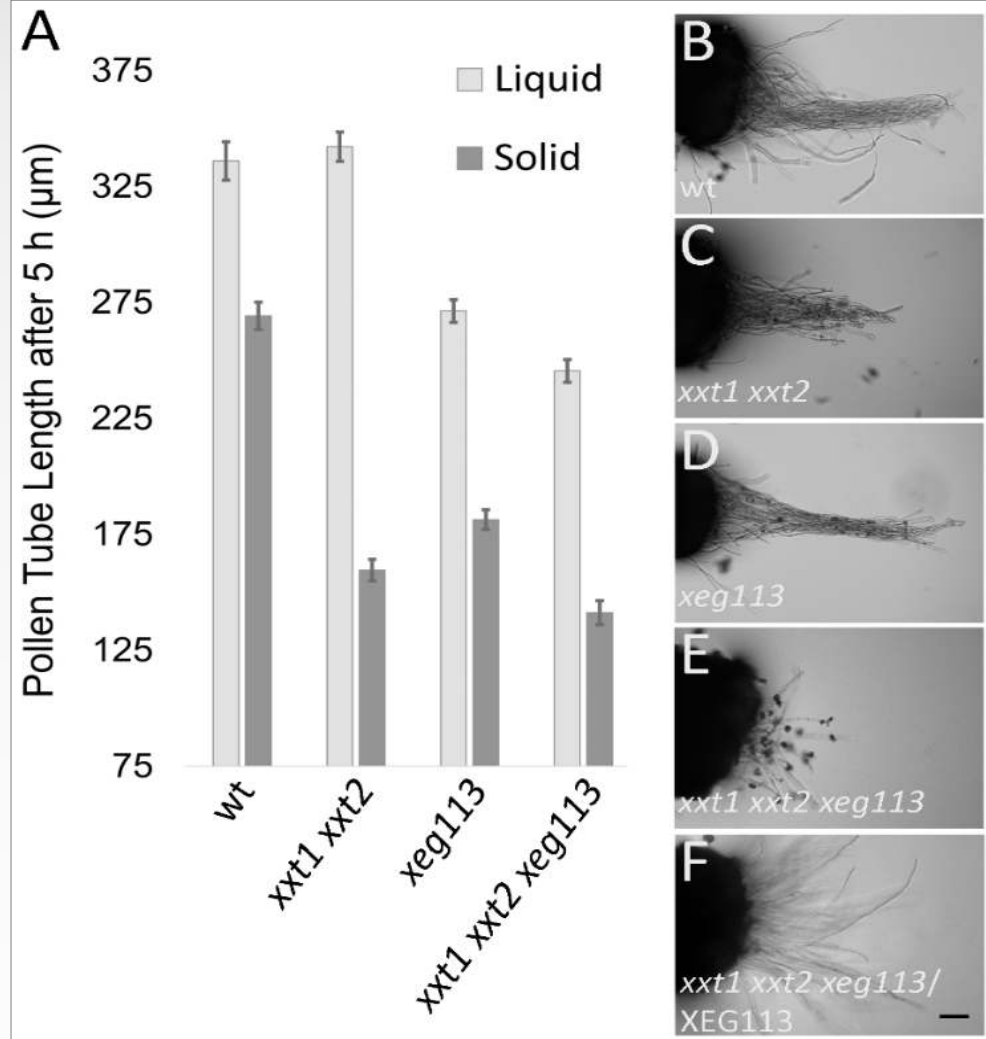
# Predicted Growth Fits Observed PT Growth Behavior



Lockhart's relation for growth in high water permeability is given by:

$$v = r_i p_i \phi$$

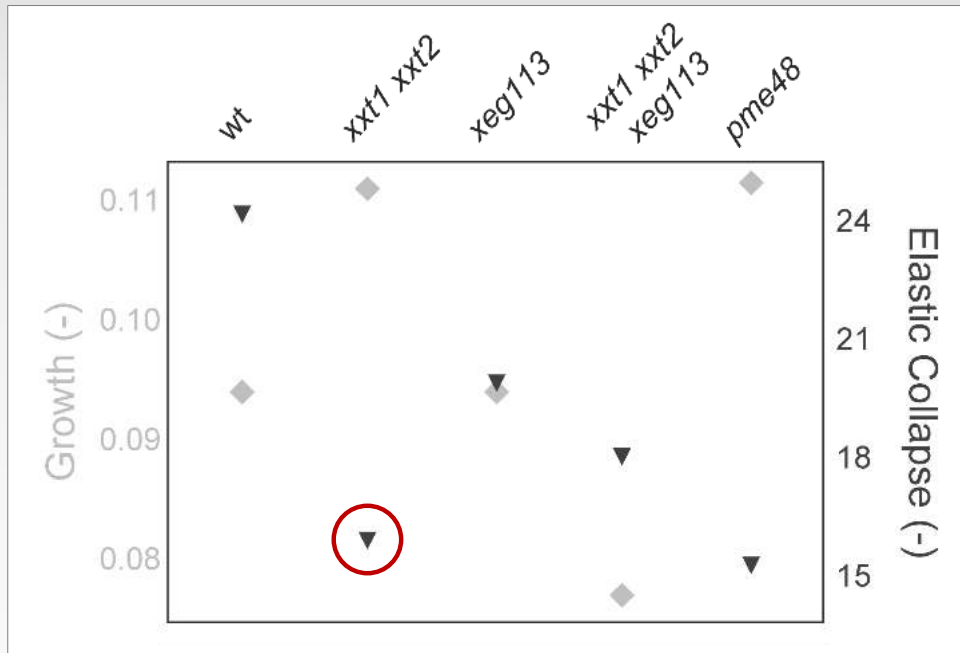
where  $v$  is the growth rate [1/hr],  $r_i$  is the radius of the cell wall not including its thickness,  $p_i$  is the turgor pressure, and  $\phi$  is the extensibility per cell wall thickness.





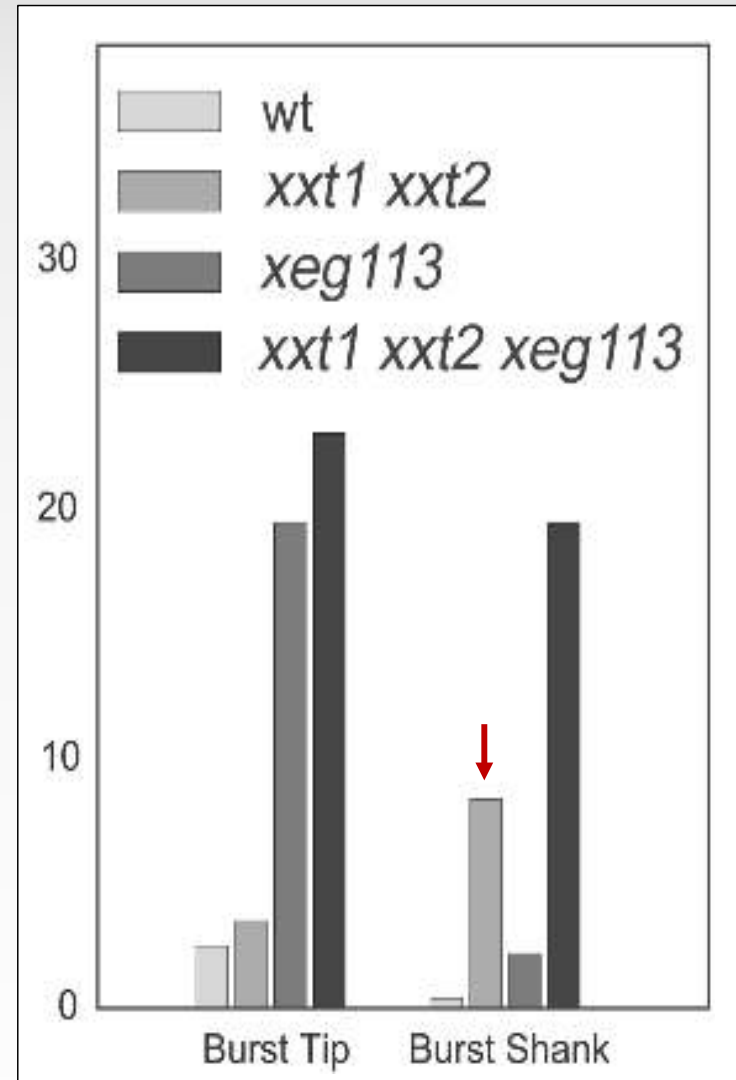
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# Shank of *xxt1 xxt2* Pollen Tubes Is Very Unstable



The ratio of the maximum von Mises stress to the critical pressure is a constant determining the proclivity of each mutant to become unstable and undergo an elastic collapse.

Simulations showed that the wild type has by far the most stable structure, followed by *xeg113* and *xxt1 xxt2 xeg113* mutants. The *xxt1 xxt2* PTs were the most unstable.



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Gautam Munglani, Tohnyui Ndinyanka Fabrice, Hannes Vogler

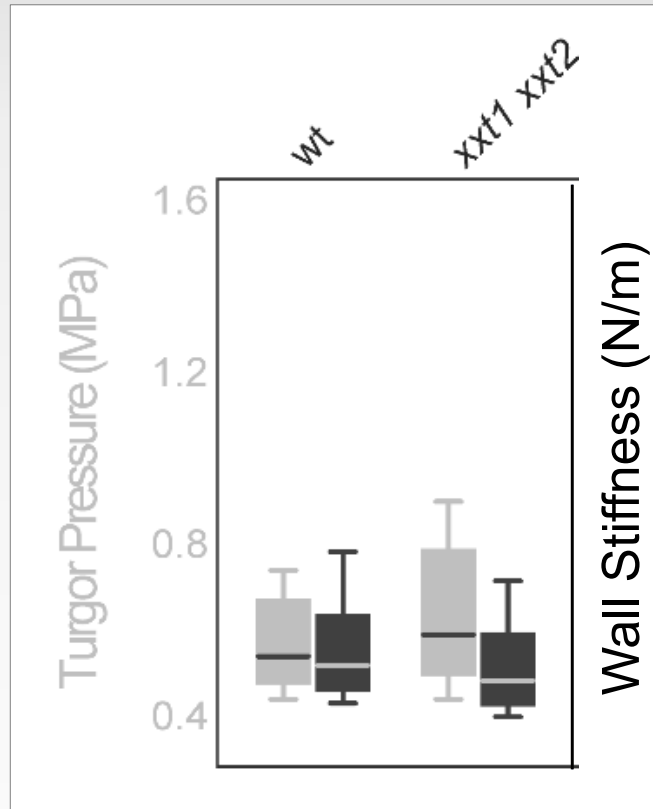
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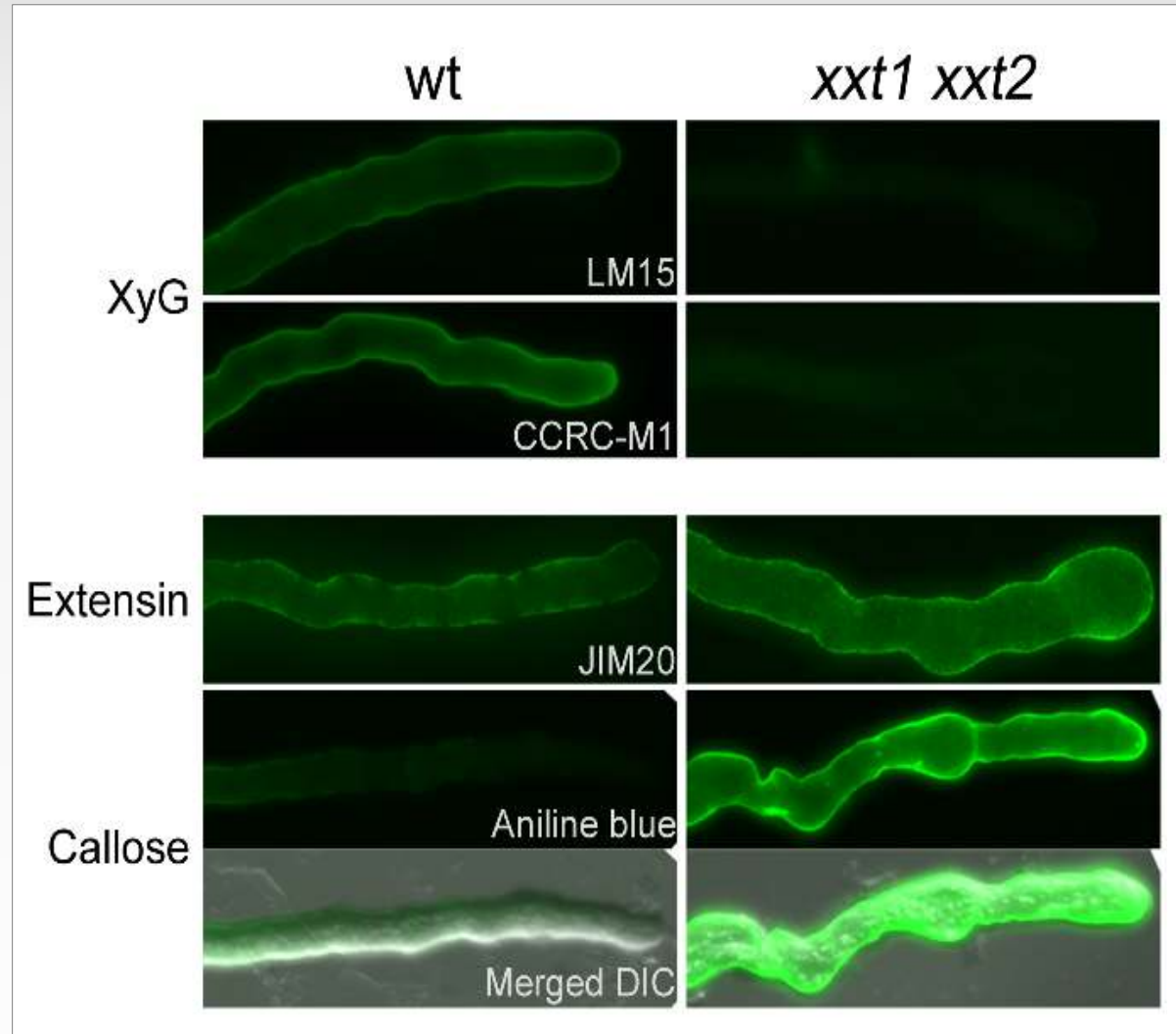


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# What Maintains the Cell Wall Stiffness in the Absence of XyG?



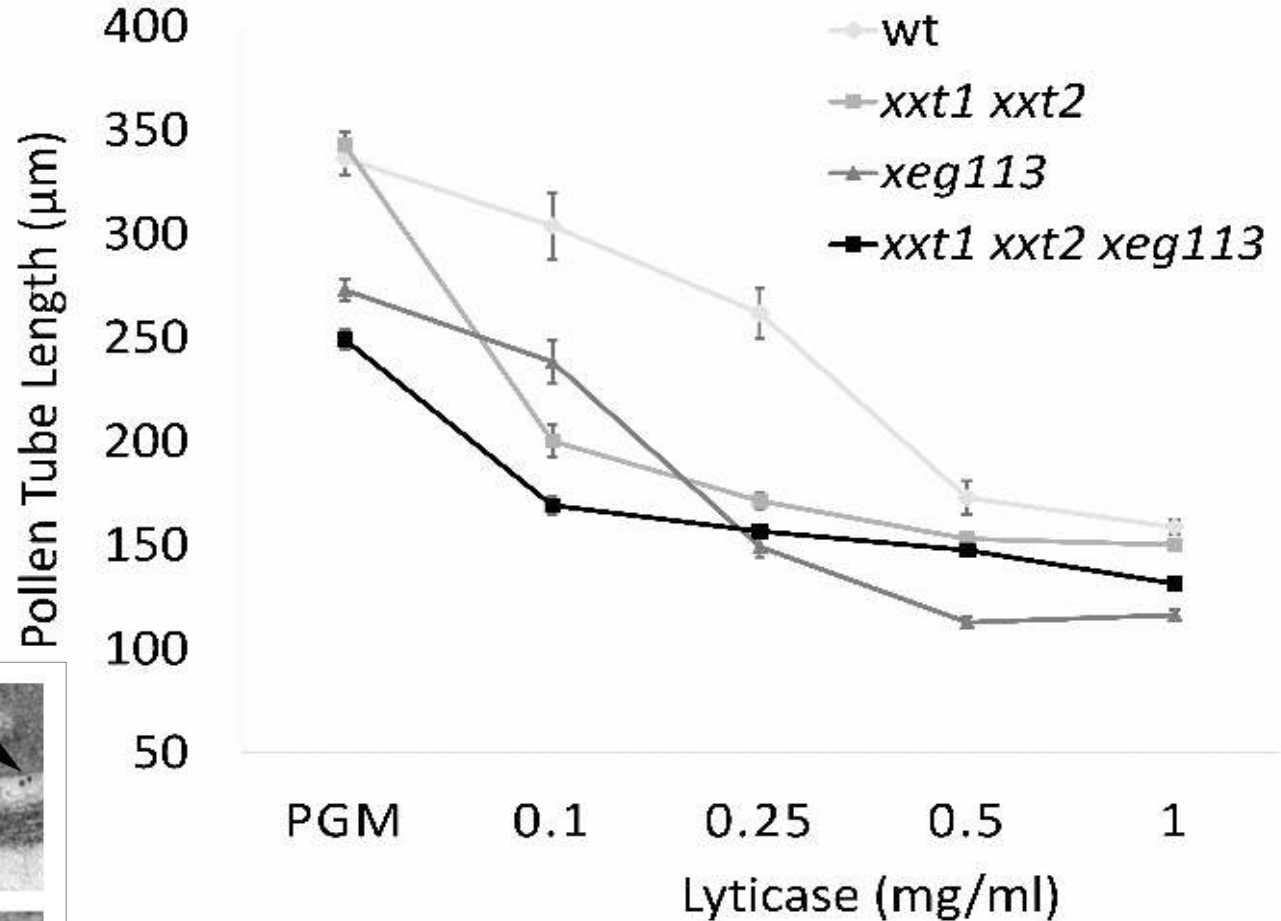
Compensatory changes: strongly increased callose deposition in XyG-deficient pollen tubes.



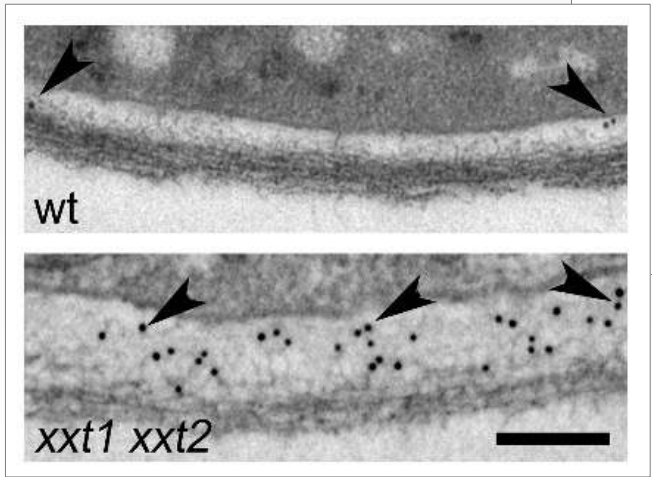


# Callose-Reinforced Pollen Tubes Are More Sensitive to Lyticase

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Immuno-Gold  $\alpha$ -Callose

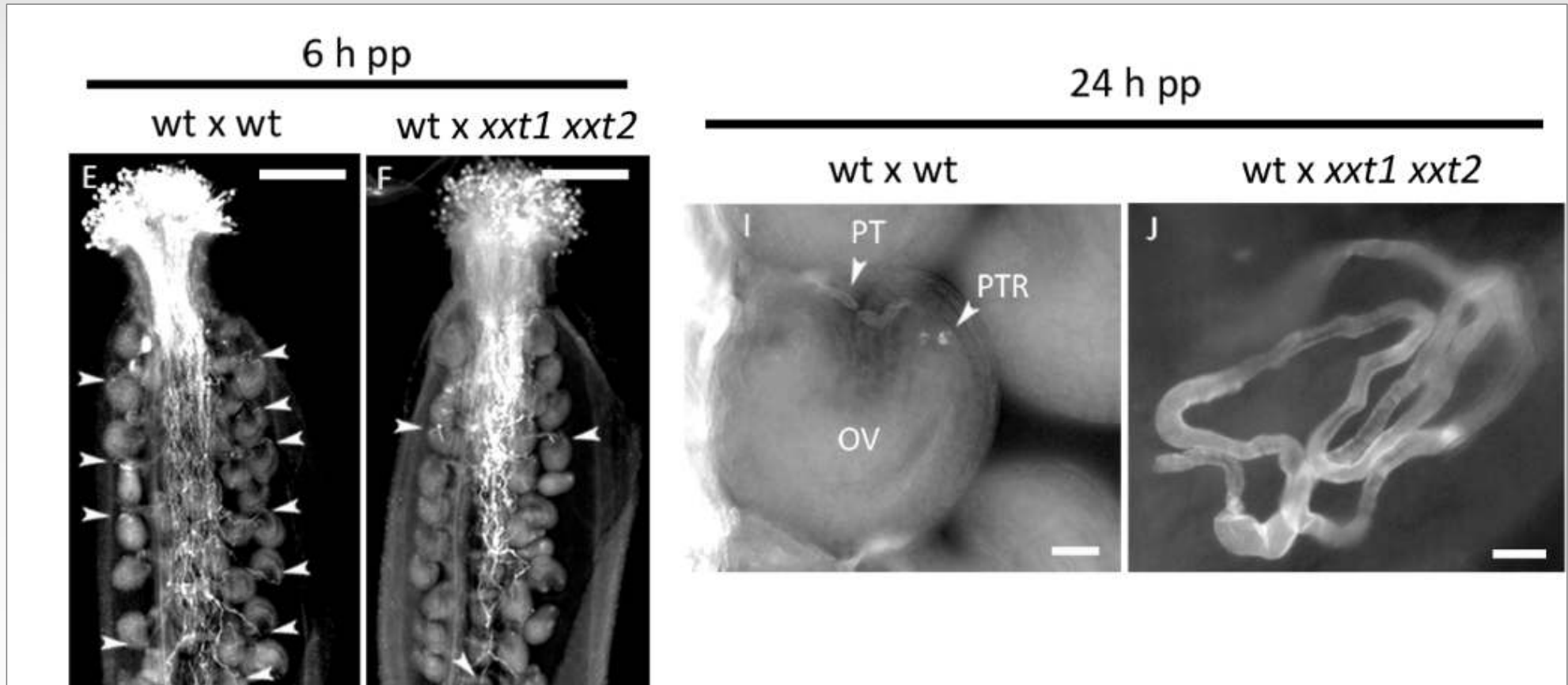






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# XyG-Deficient Pollen Tubes Grow Poorly *in vivo*

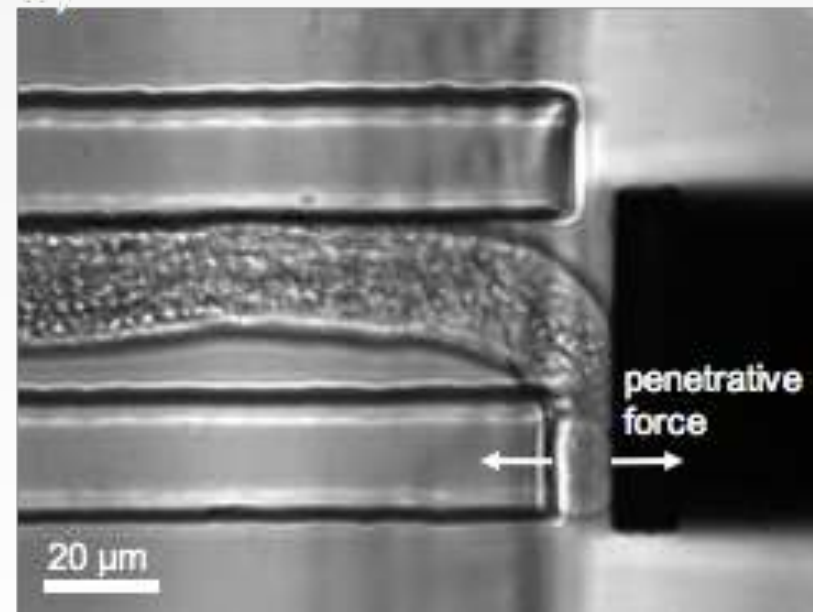
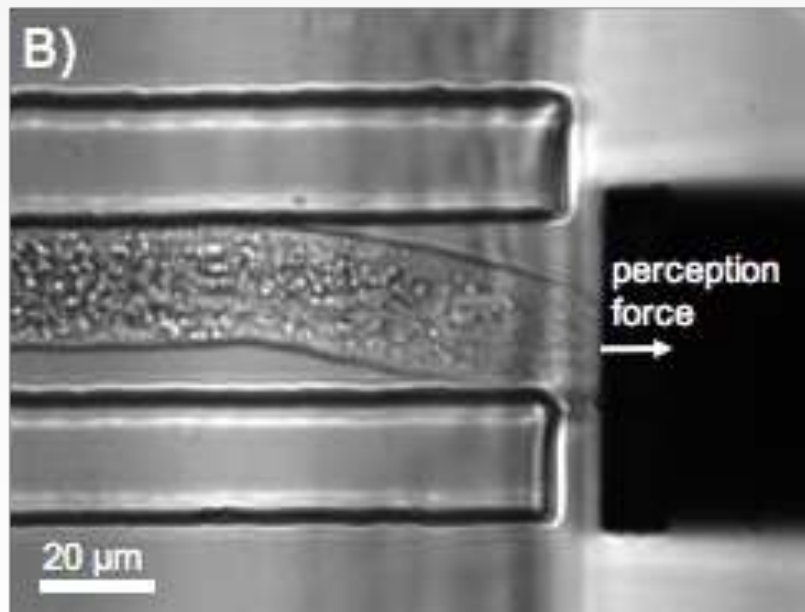
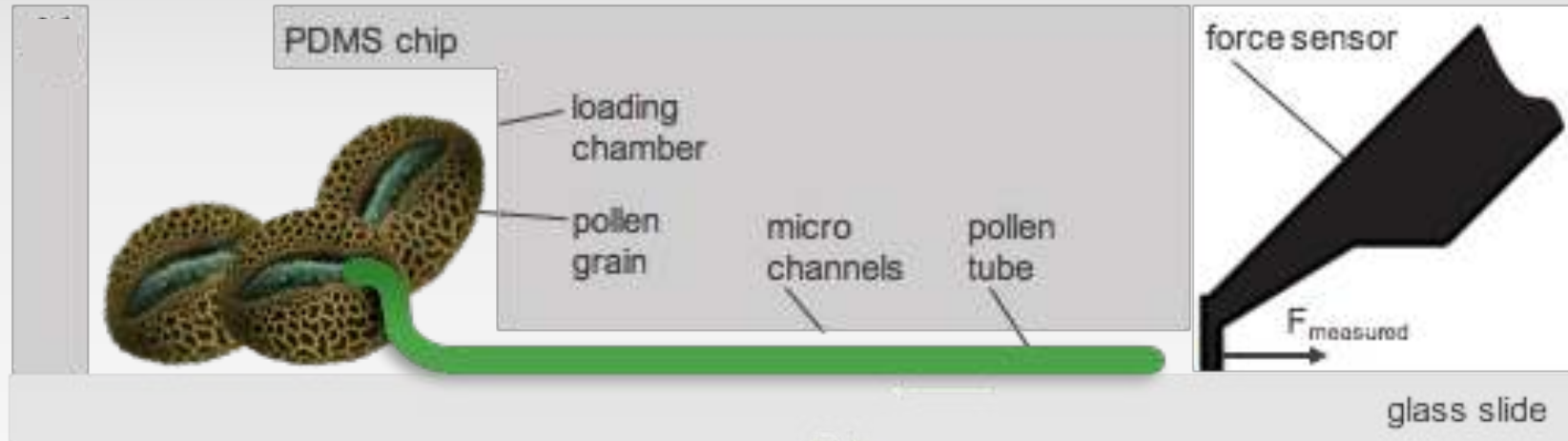


Even though *xxt1 xxt2* pollen tubes grow well *in vitro* they are poor performers *in vivo*.

Can we measure cytomechanical parameters that would approximate *in vivo* growth?



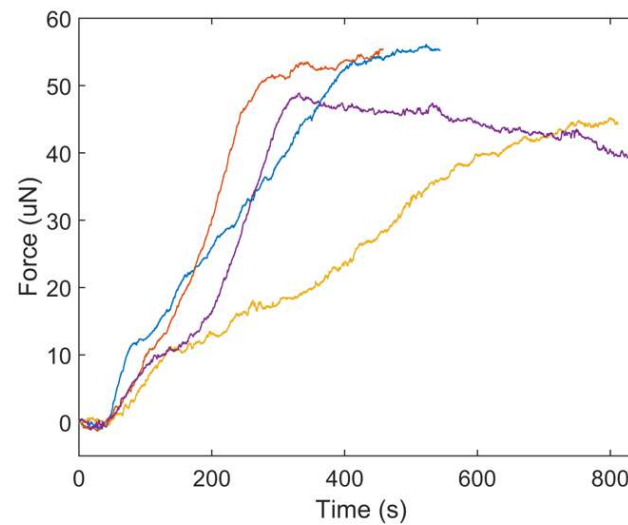
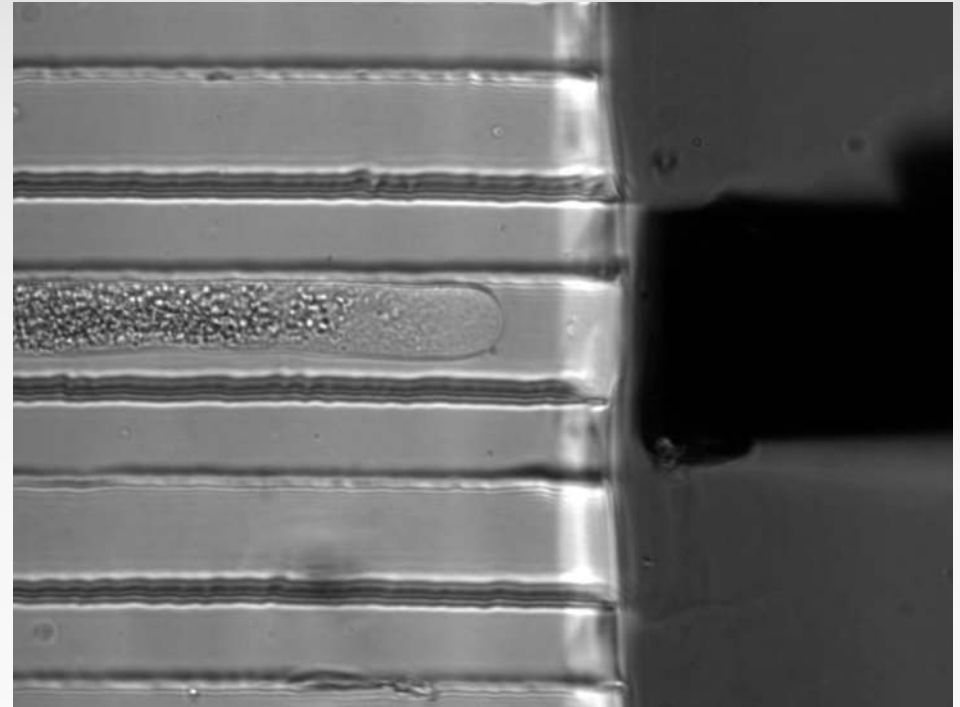
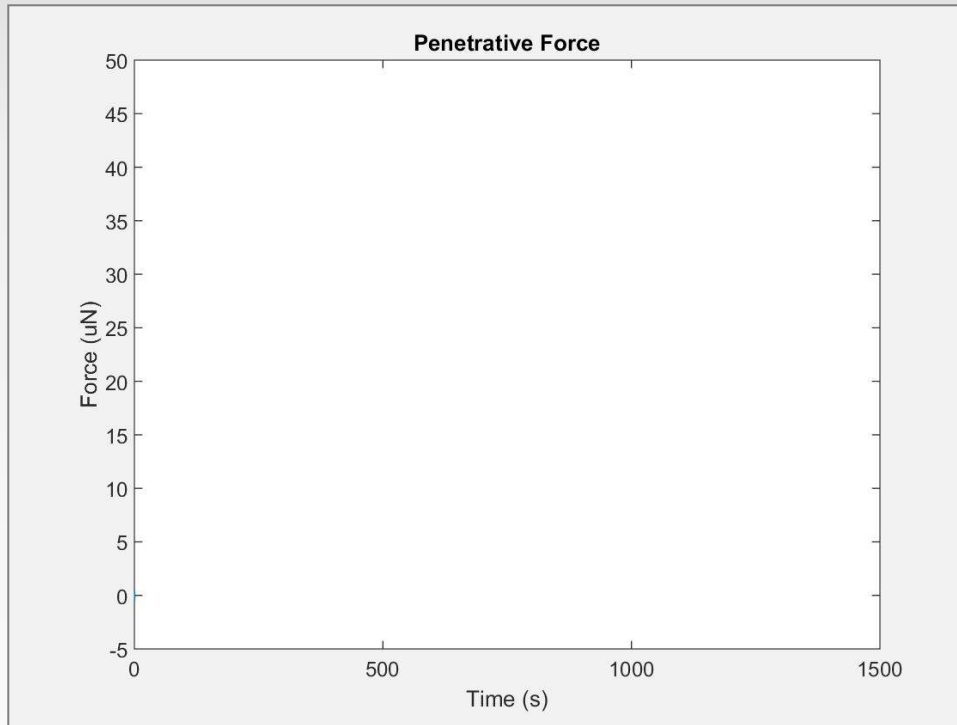
# Microfluidics Chip and Lateral Force Sensor





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# Perception and Penetrative Force Measurements



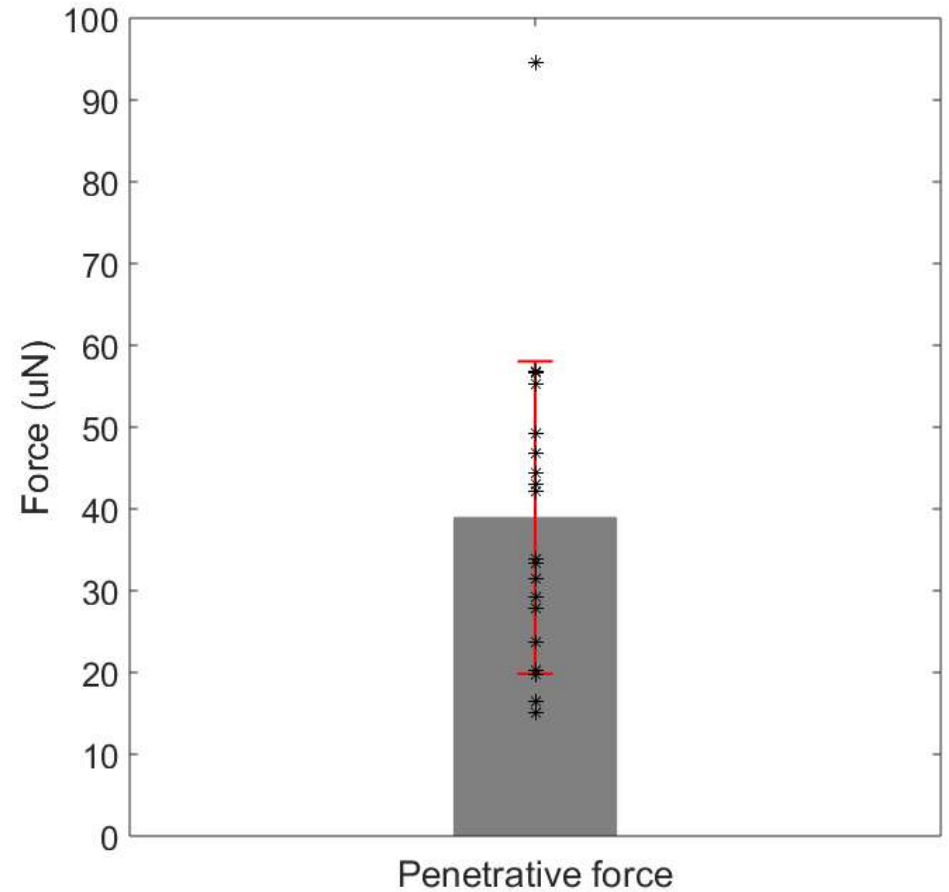
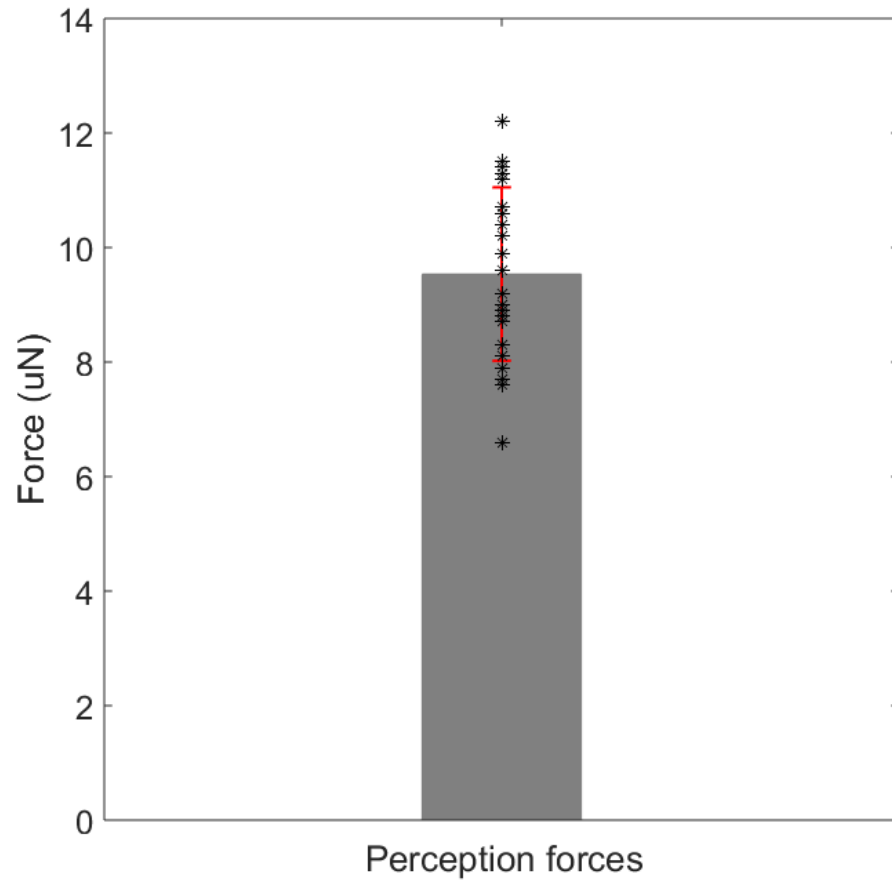
Hannes Vogler, Ian Burri





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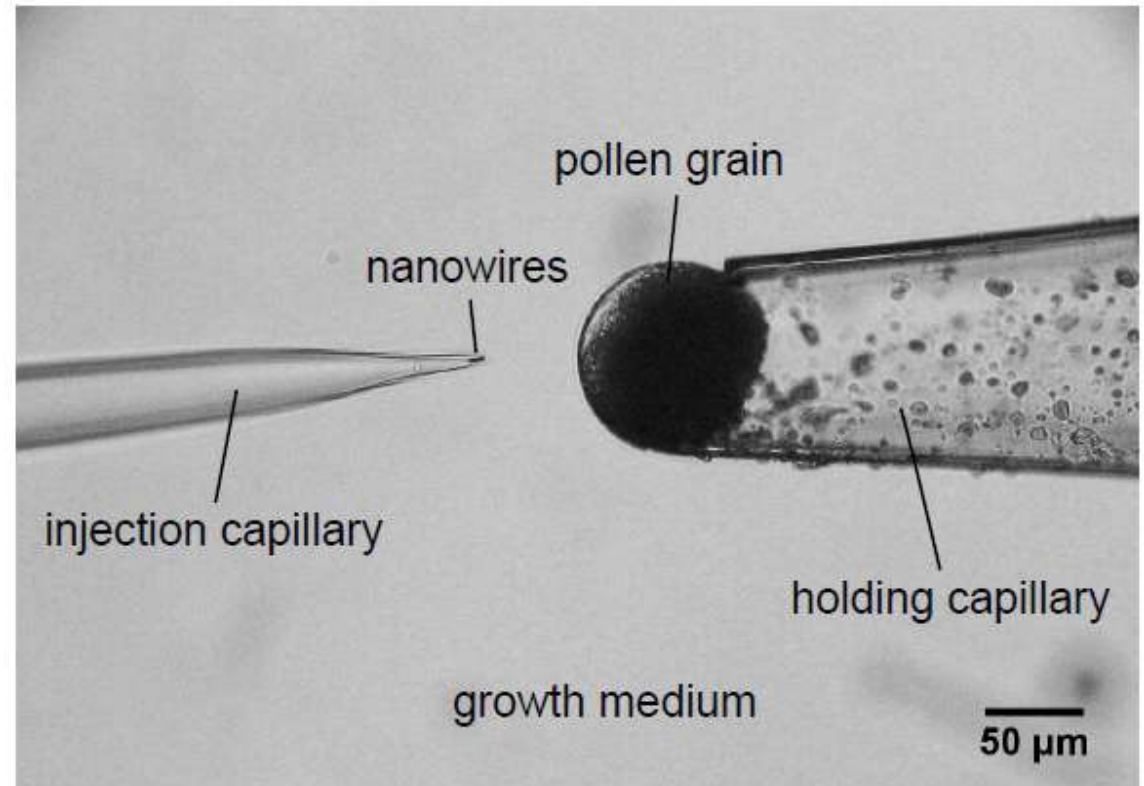
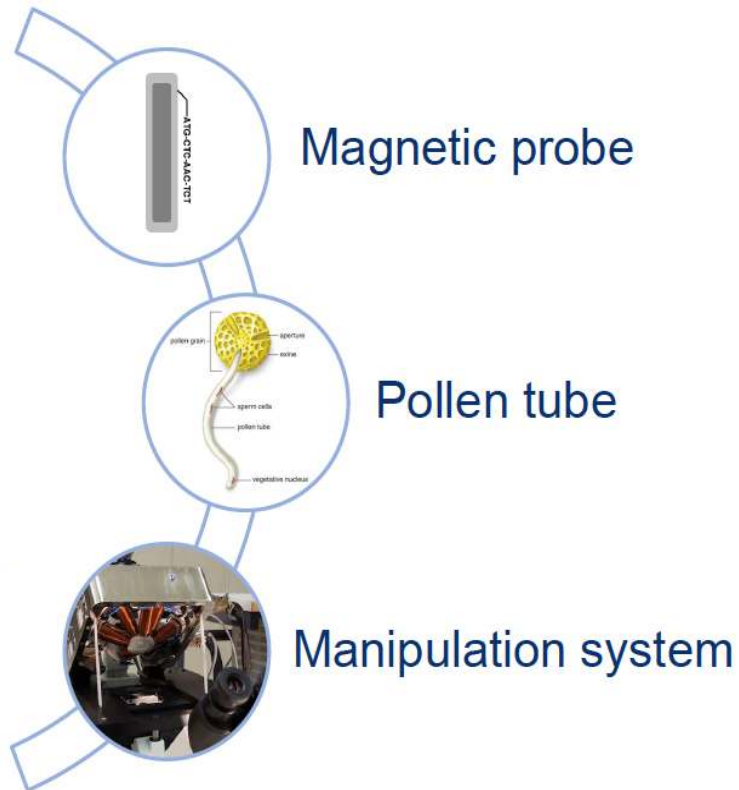
# Penetrative Forces up to 60 $\mu\text{N}$ Are Generated by Lily PTs



How does the pollen tube generate this large penetrative force?  
Rapid increase of turgor pressure?



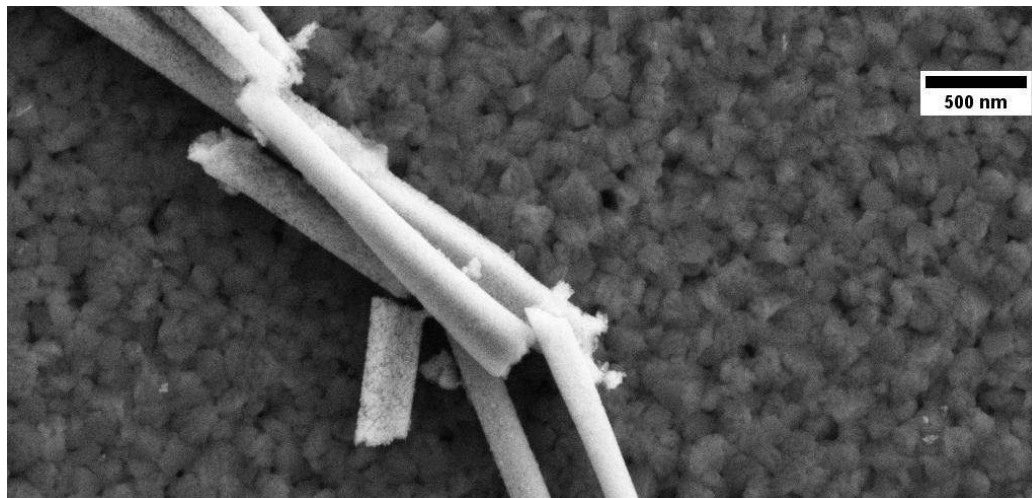
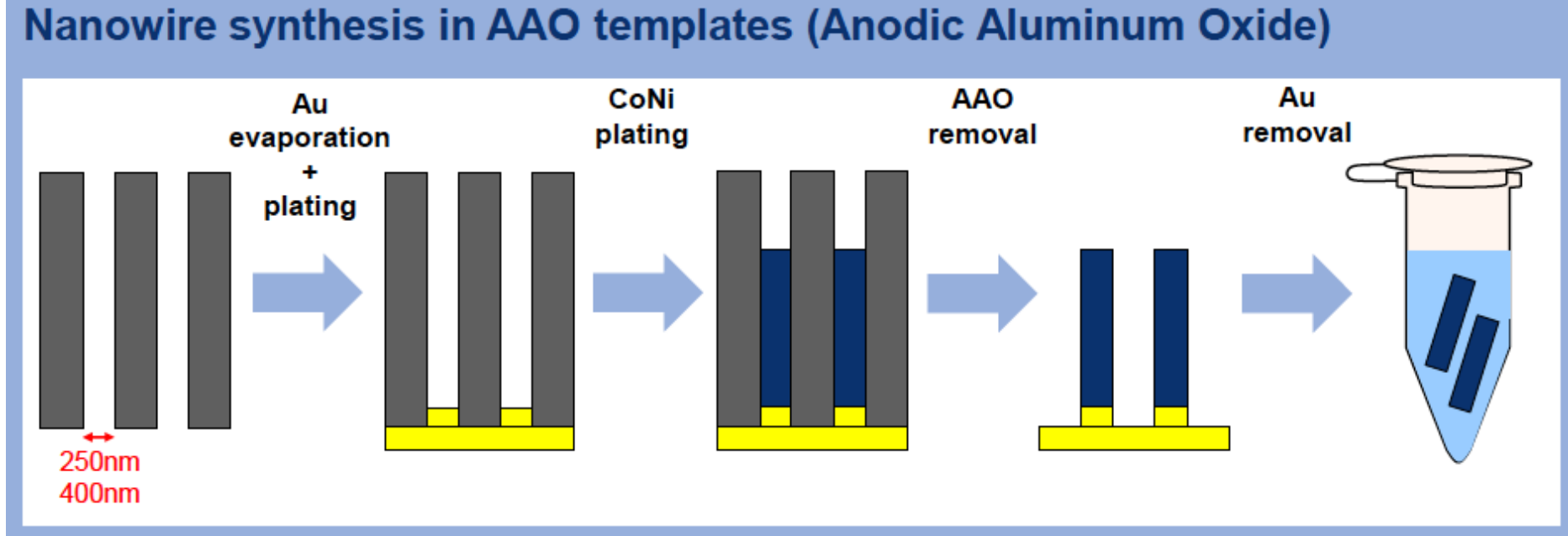
# Magnetic Nanowires to Measure Internal Biomechanical Properties



Measure and manipulate local, internal properties:

- viscosity of the cytoplasm
- cytoplasmic streaming
- cytoskeleton (local disruption)

# Nanowire Synthesis by Template-Assisted Electrodeposition

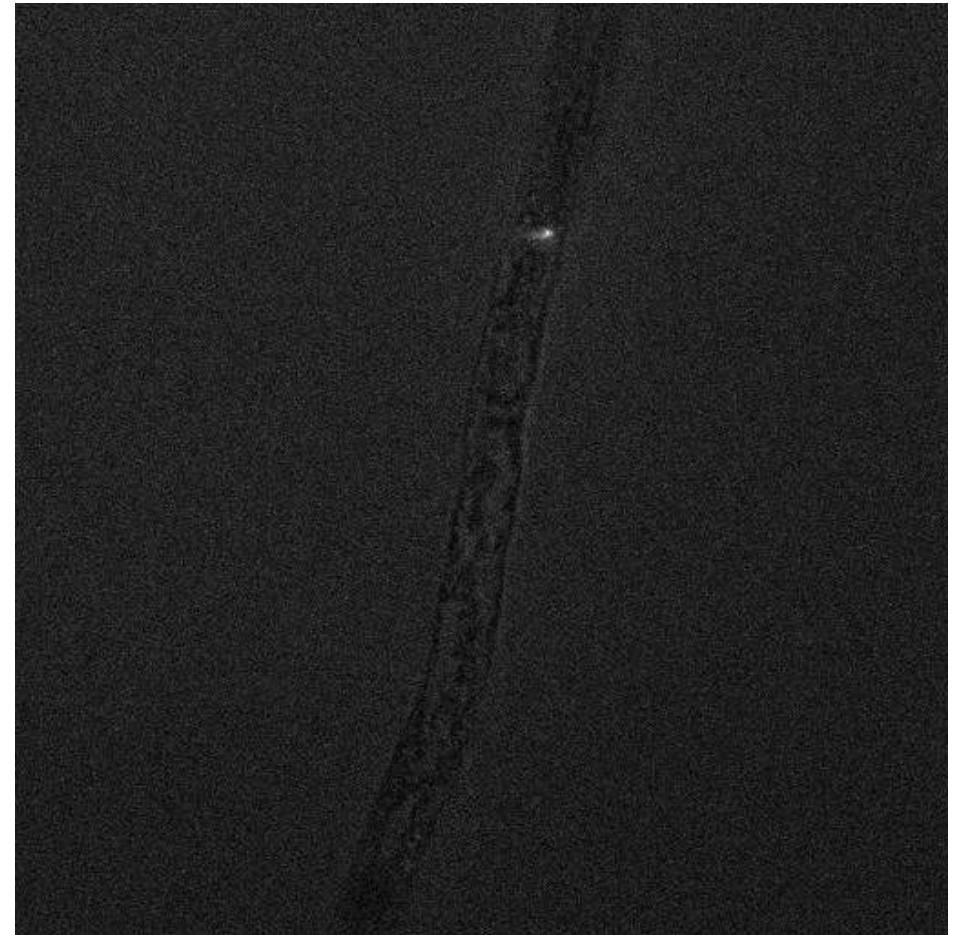


## Sol-Gel Method

To avoid agglomeration and to allow conjugation/functionalization with:

- pH-sensitive dyes (e.g. FITC)
- viscosity-sensitive dyes
- antibodies, proteins, etc.

# Injection System with Living Pollen Tube Established



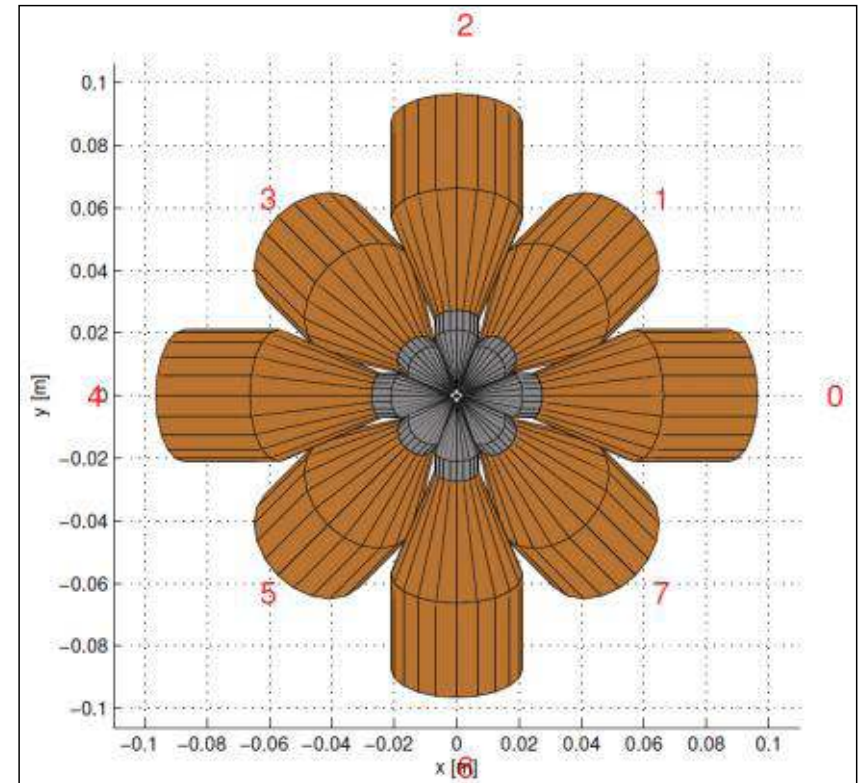
Ulrike Nienhaus, Naveen Shamsudhin

**Permanent magnet:** only rotation possible



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# Magnetic Manipulation System: *Nanomag*



**MFG-100-i (*Nanomag*):  $F_{\text{mag}} = 0.427 \text{ pN}$**

- **Gradient fields**
  - to move through the tube
- **Rotational fields**
  - to measure viscosity
- **Core extending tips**
  - to amplify magnetic field 7.5-fold



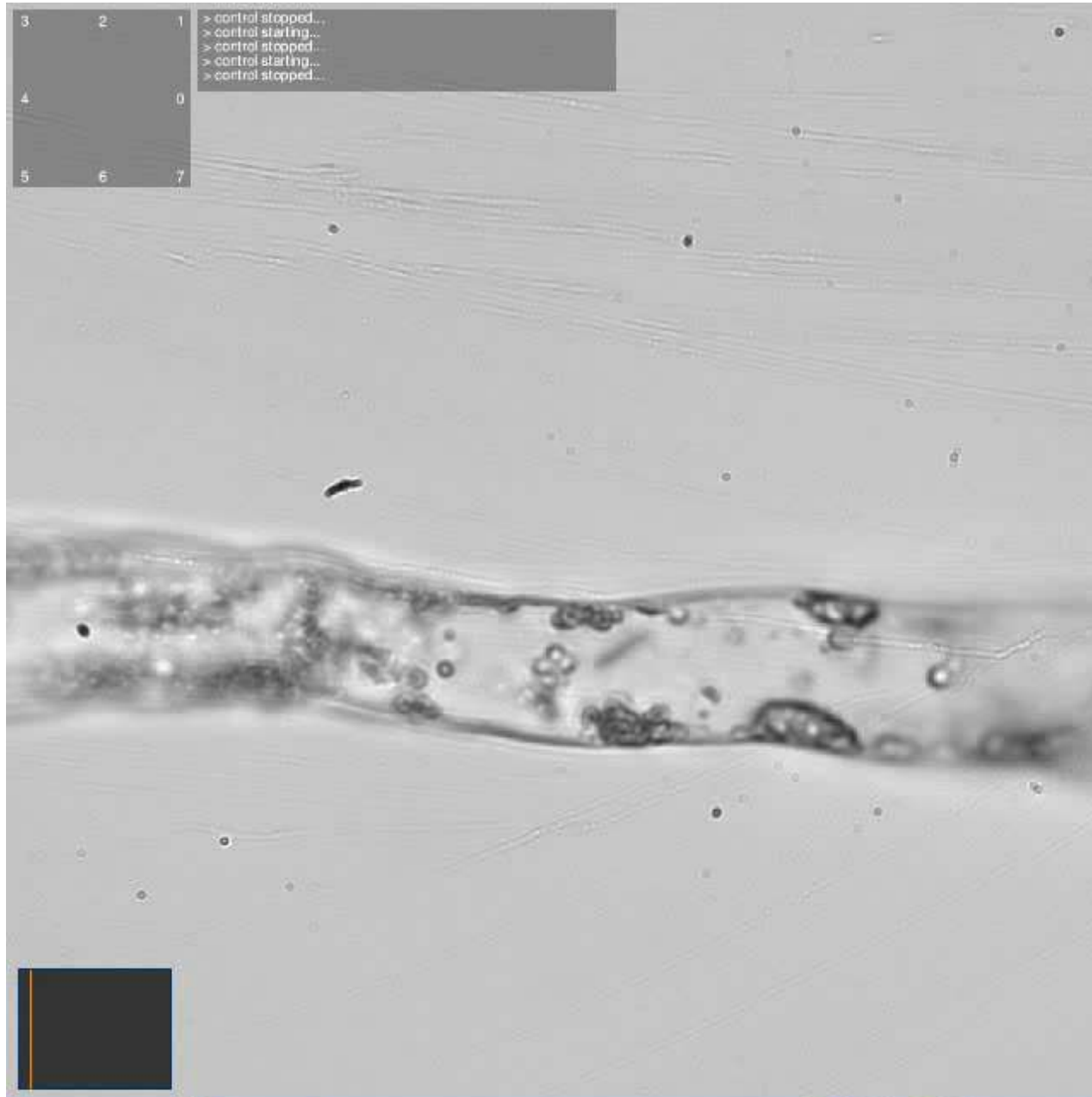
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FONDO NAZIONALE SVIZZERO  
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# Manipulations with the Nanomag



```
3 2 1  
> control stopped...  
> control starting...  
> control stopped...  
> control starting...  
> control stopped...  
4 0  
5 6 7
```



200 px

2015-05-19 12:52:01

Ulrike Nienhaus, Naveen Shamsudhin



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## Summary

- We have developed new devices to **measure mechanical properties** at the cellular level (CFM, RT-CFM, soft indentation microfluidics LoC, electrical LoC, magnetic nanowires, etc).
- Using the *Arabidopsis* pollen tube as a model, we could investigate the **relationship** between **biochemical composition** of the cell wall and its **mechanical properties**
- Interfering with cell wall composition also affects turgor pressure, revealing a tight **coordination between the cell wall and the physiological processes in the vacuole**.
- Pollen tubes are highly regulated, **fine-tuned mechanical systems** where changes in turgor pressure or mechanical cell wall properties affect growth and can lead to bursting.
- FEM/Monte Carlo-based simulations of cellular properties accurately **predict the growth behavior** of mutants with altered cell wall composition.





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**Abu Sebastian**

- Walter Haeberle

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Swiss Federal Institute of Technology Zurich

**Hans J. Herrmann**

- Gautam Munglani\*
- Falk K. Wittel

**FEMTO**  **TOOLS**

**Felix Beyeler**

- Simon Muntwyler\*

\*former members

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